



A short G1 phase is an intrinsic determinant of naïve embryonic stem cell pluripotency

Diana Coronado^{a, b, c}, Murielle Godet^{a, b, c, e}, Pierre-Yves Bourillot^{a, b, c, d}, Yann Taponnier^{a, b, c}, Agnieszka Bernat^{a, b, c, 1}, Maxime Petit^{a, b, c}, Marielle Afanassieff^{a, b, c, e}, Suzy Markossian^{a, b, c, e}, Anna Malashicheva^{a, b, c, 2}, Roberto Iacone^{f, 3}, Konstantinos Anastasiadis^f, Pierre Savatier^{a, b, c, d, e, *}

^a Stem Cell and Brain Research Institute, 69500 Bron, France

^b Inserm, U846, 18 Avenue Doyen Lépine, 69500 Bron, France

^c Université de Lyon 1, 69000 Lyon, France

^d Labex DEVweCAN

^e INRA, USC 2008, PrimaStem, 69500 Bron, France

^f BiInnovations Zentrum, Technische Universität Dresden, Tatzberg 47, 01307 Dresden, Germany

Received 10 September 2012; received in revised form 13 October 2012; accepted 13 October 2012
Available online 29 October 2012

Abstract A short G1 phase is a characteristic feature of mouse embryonic stem cells (ESCs). To determine if there is a causal relationship between G1 phase restriction and pluripotency, we made use of the Fluorescence Ubiquitination Cell Cycle Indicator (FUCCI) reporter system to FACS-sort ESCs in the different cell cycle phases. Hence, the G1 phase cells appeared to be more susceptible to differentiation, particularly when ESCs self-renewed in the naïve state of pluripotency. Transitions from ground to naïve, then from naïve to primed states of pluripotency were associated with increased durations of the G1 phase, and cyclin E-mediated alteration of the G1/S transition altered the balance between self-renewal and differentiation. LIF withdrawal resulted in a lengthening of the G1 phase in naïve ESCs, which occurred prior to the appearance of early lineage-specific markers, and could be reversed upon LIF supplementation. We concluded that the short G1 phase observed in murine ESCs was a determinant of naïve pluripotency and was partially under the control of LIF signaling.

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Abbreviations: FUCCI, Fluorescence Ubiquitination Cell Cycle Indicator.

* Corresponding author at: INSERM U846, 18 avenue Doyen Lépine, 69500 Bron, France. Fax: +33 472 01 34 61.

E-mail address: pierre.savatier@inserm.fr (P. Savatier).

¹ Present address: Laboratory of Molecular Diagnostics, Intercollegiate Faculty of Biotechnology, Medical University of Gdansk, Gdansk, Poland.

² Present address: Almazov Federal Heart, Blood and Endocrinology Centre Akkuratova, 2, 197341 Saint-Petersburg, & Department of Embryology, St. Petersburg State University, Universitetskaya nab., 7/9, St. Petersburg, 199034, Russia.

³ Present address: F. Hoffmann-La Roche Ltd., Pharma Research and Early Development, Basel, CVM Discovery, Switzerland.

Introduction

Mouse embryonic stem cells (ESCs) are derived from the early epiblast of a pre-implantation blastocyst. They can self-renew in the so-called naïve state of pluripotency by exploiting LIF and BMP4 signaling pathways to block differentiation (Ying et al., 2003). It has been found that blockade of both extracellular regulated kinase (ERK) and glycogen synthase kinase-3 (Gsk3) pathways with selective small molecule inhibitors is sufficient to stabilize and sustain ESC self-renewal in the so-called ground state of pluripotency (Ying et al., 2008). Upon withdrawal of LIF and activation of FGF2 signaling, ESCs acquire the characteristics of Epiblast Stem Cells (EpiSCs) and self-renew in the so-called primed state of pluripotency (Brons et al., 2007; Tesar et al., 2007; Takehara et al., 2011). Ground pluripotency is believed to match the status of the pluripotent stem cells of the inner cell mass (ICM), whereas primed pluripotency may reflect the status of the late epiblast cells of the post-implantation embryo (Nichols and Smith, 2009).

ESCs in the naïve state of pluripotency are rapidly proliferating mammalian cells that exhibit an unusual cell cycle structure characterized by a G1 phase of less than 2 h (Savatier et al., 1994). This short duration is associated with a unique mechanism of cell cycle regulation, which is highlighted by lack of MAPK, cyclin D, and pRB control (Jirmanova et al., 2002; Savatier et al., 1996; White et al., 2005), lack of a functional p53–p21 pathway in response to DNA damage (Aladjem et al., 1998), active transcription of E2F target genes, and robust activity of cyclin E-Cdk2 and cyclin A-Cdk2 complexes throughout the cell cycle (Stead et al., 2002; White et al., 2005). As ESCs differentiate, their cell cycle structure drastically changes. A longer duration of the G1 phase results, and the mechanism of cell cycle regulation changes to that typically observed in other mammalian cells including the acquisition of p53- and RB-dependent checkpoints. The reason for the short G1 phase exhibited by ESCs is unclear. It might simply reflect a fundamental requirement of rapid proliferation of epiblast cells to initiate gastrulation in the developing embryo. However, it is possible that a short G1 phase might be involved in actively sustaining the pluripotent state, as discussed previously (Burdon et al., 2002; Kapinas et al., 2013).

An equally important question is the role of LIF signaling in regulating the ESC cell cycle. Some target genes of the LIF/STAT3 pathway, such as *c-myc*, *Klf4*, *Klf5*, and *Pim-1* (Aksoy et al., 2007; Bourillot et al., 2009), participate in regulating the G1-to-S phase transition in differentiated cells and are often deregulated in cancer cells (Bureau et al., 2009; Ghaleb et al., 2005; Shirogane et al., 1999). In mESCs, *Klf5* activates the G1/S transition by promoting Akt phosphorylation and inhibiting the kinase inhibitor p21cip1 (Ema et al., 2008). Thus, it remains to be determined whether LIF signaling contributes to the regulation of the G1-to-S phase transition in naïve ESCs.

The Fluorescence Ubiquitination Cell Cycle Indicator (FUCCI) reporter system was developed to monitor cell cycle progression in live cells (Sakaue-Sawano et al., 2008). Using this reporter system, we uncovered the alterations in the cell cycle distributions associated with the transitions between the ground, naïve, and primed pluripotent states. We explored the cell cycle dependency of differentiation commitment in

naïve ESCs and the role of cyclin E in this commitment. We also uncovered the role of LIF in regulating the G1-to-S phase transition.

Material and methods

Plasmid construction and shRNA design

pFUCCI G1 orange and pFUCCI S/G2/M green plasmids (MBL International Corporation, ref AM-VS0605) were digested with XbaI and BamHI. The resulting 1045-bp and 1106-bp fragments containing mKO2:Cdt1(30:120) and hmAG1:Geminin (1:110) coding sequences, respectively, were sub-cloned between the HindIII and BamHI sites in *pGAE-CAG-eGFP-WPRE* (Wianny et al., 2008) to generate *pGAE-mKO2:Cdt1* and *pGAE-mAG:Geminin*, respectively.

Expression vectors for rat and human cyclin E that were suitable for conditional expression induced by Dox and Dex (Anastassiadis et al., 2002) were generated. Myc-tagged rat cyclin E cDNAs (both WT and S180D mutant) were amplified by PCR from *pINDCycE^{WT}* and *pINDCycE^{S180D}*, respectively (Matsumoto and Maller, 2004), using primers containing Sall and NotI sites (underlined) (Myc-cycE-Sall-F: 5'-ACGCGTCGAC ACCATGGAGCAAAGCTCATTTTC-3'; Myc-cycE-NotI-R: 5'-AAGG AAAAAAGCGGCCGCTCATTCTGTCTCTCTGCTCAC-3'). Human cyclin E cDNA was amplified by PCR from *pCS2cycE^{1WT}* (Geng et al., 2007) using the following primers: hCycE1SallF: 5'-ACG CGTCGACACCATGCGGAGGGAGCGCAGGGAC-3'; hCycE1Not1R: 5'-AAGGAAAAAGCGGCCGCTCACGCCATTTCCGGCCCCGC-3'. The PCR products were subsequently subcloned between the Sall and NotI sites in *pSport1-SV40-PolyA* to generate *pSport1-cDNA* plasmids. Similarly, *pTet/CMV-INS2-Hygro* was digested with Sall and SwaI. The resulting 5046-bp *Tet/CMV-INS2-Hygro* fragment was subcloned into each *pSport1-cDNA* plasmid by Red/ET recombination to generate *pSport1-tet/CMV-INS2-cDNA-Hygro* vectors.

Cyclin E (CycE) shRNA sequences were designed using the siDESIGN®Center application of Dharmacon (<http://www.dharmacon.com>). shRNA sequences were cloned into *pENTRY* and tested by transient transfection in CGR8 ESCs. The sequences of the tested shRNAs and interference resulting from transfection of *pENTRY-shRNA* vectors are given in Supplementary data 1. The selected shRNA sequence and the control scrambled shRNA were subcloned into pLenti6/*BLOCK-iT-PGKneo^r* (Bourillot et al., 2009).

ESC culture, conversion into cEpiSCs, LIF rescue assay and differentiation

CGR8 and the E14tg2a-derivative *Rex1^{GFP-bla}/Oct4^{CFP-puro}* ESC lines (129/Sv strain) were routinely cultured in the absence of feeder cells in Glasgow's Modified Eagle's Medium (GMEM) supplemented with 10% fetal calf serum (PerbioScience CRC0406) and 1000 U/ml of LIF (Savatier et al., 1994; Toyooka et al., 2008). They were also cultured in N2B27 medium supplemented with PD0325901, CHIR99021, and 1000 U/ml LIF (2i/LIF medium). To convert ESCs to cEpiSCs, culture dishes were pre-coated with 15 mg/ml of human fibronectin (Chemicon) for 2 h at 37 °C. Subsequently, 10⁶ ESCs were plated in chemically defined medium (Brons et al., 2007) supplemented with Activin A (10 ng/ml; R&D systems) and

FGF2 (12 ng/ml; R&D systems). After 1 week, cells were treated with 0.1% trypsin-EDTA and the single cell suspension was plated onto a new pre-coated plate. After 1 week, cells were harvested using 0.5 mg/ml collagenase II (Sigma) and plated onto two new plates. During the next 3 weeks, converted ES cells were passaged every 3 days with collagenase treatment at a subculture ratio of 1:3. cEpiSC cell lines were considered to have been established after at least seven passages.

For LIF rescue assays, ES cells were plated at 10^3 cells per gelatin-coated 100-mm tissue culture dish in complete ES cell medium. Cells were exposed to medium without LIF for 12–48 h and subsequently cultured in normal ES cell medium with 1000 U/ml of LIF for 6 days. To detect cells that expressed alkaline phosphatase, dishes were fixed in methanol for 15 min, followed by staining for 15 min with a solution containing 1 mg/ml Fast Red TR saltTM (Sigma) dissolved in 0.1 M Tris, pH 9.2, containing 200 mg/ml naphthol AS-MX phosphate. For each treatment, the percentages of undifferentiated, mixed, and differentiated colonies were determined in triplicate.

To induce differentiation, cells were allowed to aggregate in hanging drops in ES cell medium without LIF (100 cells/drop). After 3 days, EBs were collected and then grown in suspension for 1–7 days in non-adherent Petri dishes.

Production of lentiviral vectors and infection of ESCs

To produce SIV-derived lentivectors, 293 T cells were transfected with a mixture of DNA containing 7.5 μ g of a *pGRev* plasmid encoding for the vesicular stomatitis virus glycoprotein (VSV-G) envelope; 4 μ g of a *pSIV3+* plasmid encoding for the gag, pol, tat, and rev proteins; and 11.5 μ g of vector plasmids using the calcium phosphate precipitation technique. For infection, CGR8 and *Rex1^{GFP-bla}/Oct4^{CFP-puro}* ESCs were plated at a density of 10^4 cells in 24-well plates in 1 ml of medium containing 100 μ l of ES cell medium and 900 μ l of culture supernatant from virus producer cells. After 48 h, ES cells were trypsinized, replated at 10^4 cells per gelatin-coated 10 mm tissue culture dish, and cultured in complete ES cell medium.

Semi-quantitative and real-time PCR analyses

Total RNA was extracted using RNeasy kits with on-column DNase digestion (Qiagen). Reverse transcription was performed using M-MLV Reverse Transcriptase (Promega Corp., Madison, WI) and Random Primers (MWG-Biotech AG, Ebersberg, Germany) according to the manufacturer's recommendations. PCR reactions were performed with a Perkin-Elmer thermal cycler at 94 °C for 30 s, 55–62 °C (according to primers) for 30 s, and 72 °C for 60 s. Quantitative PCR was performed with Step One Plus thermal cycler (Applied Biosystem) at 94 °C for 45 s, 55 °C for 45 s, and 72 °C for 45 s for 40 cycles with Quantitec SYBR Green (Qiagen). In all cases, the expressions of the target genes were normalized to that of *β -actin*. Primers, annealing temperatures, and number of cycles were as described in a previous study (Bourillot et al., 2009). Primer sequences for mouse/rat cyclin E (mCycE) were 5'-TGGAG TTGATGCAGAAGTC and

3'-ATGGCTTCTTTGCTTGGGC. Primer sequences for human cyclin E (hCycE) were 5'-AGCACTTCAGGGGCGTCGC and 3'-CTGGGGAGAGGAGAAGCCC.

Flow cytometry and fluorescence microscopy

Cells were washed with PBS and dissociated by treatment with trypsin-EDTA. For cell cycle analysis, $1 \cdot 10^6$ cells were incubated with 20 μ l of Hoechst 33342 (SIGMA B2261) in the dark for 30 min at 37 °C. For SSEA1 immunostaining, $5 \cdot 10^5$ cells were incubated for 15 min at room temperature in GMEM+10% fetal calf serum. Then 10 to 30 ng of SSEA1-Alexafluor 647 (sc21702, Santa Cruz) antibody was added and cells were further incubated for 30 min at room temperature. Cells were analyzed using a FACS LSR II (Becton-Dickinson) equipped with 355, 488, and 561 nm lasers or were sorted with an Aria cell sorter (Becton-Dickinson) equipped with 405, 488, and 633 nm lasers. Data were acquired and analyzed using DiVa software.

Time-lapse videomicroscopy recording

Cells were grown in a 5% CO₂ atmosphere at 37 °C for 24 h in a Pecon incubating chamber placed on a Leica DMIRBE inverted microscope stage. Observations were made with a 20 \times objective under halogen illumination and fields were scanned using Metamorph software.

Results

Direct visualization of cell cycle progression using the FUCCI reporter

The Fluorescence Ubiquitination Cell Cycle Indicator (FUCCI) reporter system uses two distinct fluorescence-emitting proteins to distinguish cells in the G1 phase from those in the other cell cycle phases (Sakaue-Sawano et al., 2008). Thus, it provides for direct visualization of cell cycle progression and FACS sorting. We engineered an ESC line that stably expressed FUCCI reporters. For this purpose, mouse CGR8 ESCs were transduced with two lentiviral vectors that expressed orange-emitting Cdt1-mKO2 and green-emitting Geminin-mAG fusion proteins, both of which were under the control of the robust CAG promoter. One clone (designated ESC-FUCCI) that expressed both fluorescent proteins at high levels was chosen for subsequent studies (Fig. 1A). Flow cytometry analysis for orange and green fluorescence, which was associated with DNA staining with Hoechst, demonstrated the following: (i) 10.4% of ESC-FUCCI cells had a 2n DNA content and no fluorescent signals above background [mKO2(-)mAG(-)], which corresponded to cells in early G1 phase (Fig. 1B); (ii) 9.4% had a 2n DNA content and exhibited an orange fluorescent signal [mKO2(+)mAG(-)], which corresponded to cells progressing toward the G1/S transition; and (iii) 74.8% had a >2n DNA content and expressed a green fluorescent signal [mKO2(-)mAG(+)]. mAG-positive cells that exhibited low green fluorescence intensity were primarily in the S phase [designated (mKO2(-)mAG(+)) cells], whereas those with strong green fluorescence intensity were primarily in the G2 phase [designated (mKO2(-)mAG(++)) cells]. LIF deprivation for 36 h dramatically increased the percentage of [mKO2(+)mAG(-)]

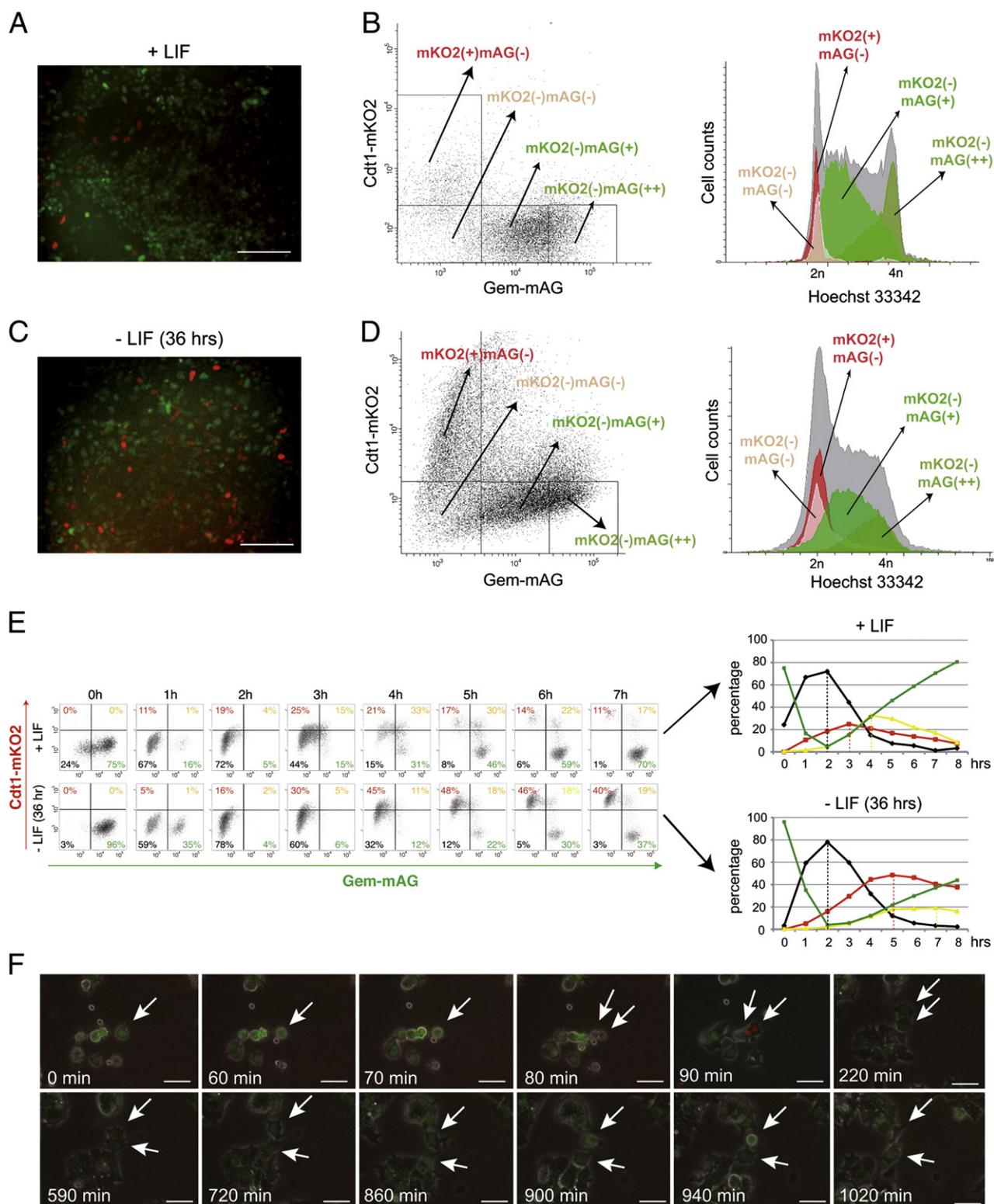


Figure 1 Generation of pluripotent stem cell lines expressing the FUCCI reporter. (A, C) Representative fluorescence images of ESCs expressing mKO2-hCdt1 and mAG-hGem in the presence of LIF (A) or after LIF withdrawal (C) for 36 h. Scale bar represents 100 μ M. (B, D) Dot plots of ESC-FUCCI (+LIF) and LIF-deprived ESCs (-LIF 36 h) showing red [mKO2(+)/mAG(-)], yellow [mKO2(+)/mAG(+)], and green [mKO2(-)/mAG(+)] fluorescence. The histograms show the cell cycle position of each cell type after staining with Hoechst 33342 and FACS analysis. (E) Synchronization of ESC-Fucci with colcemid (60 ng/ml for 4 h) followed by their release from the mitotic block. Cells were harvested at different time points and the percentages of [mKO2(-)/mAG(-)], [mKO2(+)/mAG(-)], [mKO2(+)/mAG(+)], and [mKO2(-)/mAG(+)] cells were analyzed by flow cytometry. (F) Cell cycle-dependent changes in fluorescence of mKO2-hCdt1 and mAG-hGem in ESC analyzed by time-lapse videomicroscopy. Arrows indicate the tracked cells. Scale bar represents 10 μ m.

cells, which reflects the lengthening of the G1 phase that accompanies ESC differentiation (Savatier et al., 1994) (Figs. 1C,D).

ESC–FUCCI were synchronized by colcemid treatment for 5 h followed by mitotic shake-off. Between 1 and 8 h after their release from the mitotic block, four successive waves of cells were observed (Fig. 1E): (i) a first wave of [mKO2(–) mAG(–)] cells (early G1); (ii) a second wave of [mKO2(+) mAG(–)] cells (late G1); (iii) a third wave of [mKO2(+) mAG(+)] cells (G1/S transition); and (iv) a fourth wave of [mKO2(–) mAG(+)] cells (S and G2 phases). These observations were consistent with the rapid transition from the G1-to-S phase that characterizes ESCs (Savatier et al., 1994). In a parallel experiment, ESC–FUCCI were deprived of LIF for 36 h prior to synchronization. The first wave of [mKO2(–) mAG(–)] cells peaked at 2 h, as previously observed with LIF-stimulated cells. In contrast, the waves of [mKO2(+) mAG(–)], [mKO2(+) mAG(+)], and [mKO2(–) mAG(+)] cells were delayed, which indicated that LIF withdrawal increased the G1 phase duration and delayed the entry of cells into the S phase. A cell cycle study using time-lapse videomicroscopy revealed transitions from [mKO2(–) mAG(++)] to [mKO2(–) mAG(–)], from [mKO2(–) mAG(–)] to [mKO2(+) mAG(–)], from [mKO2(+) mAG(–)] to [mKO2(+) mAG(+)], and from [mKO2(+) mAG(+)] to [mKO2(–) mAG(+)]. This confirmed the identity of each colored phenotype (Fig. 1F).

G1 phase duration varies according to the pluripotency state

Mouse ESCs display a mosaic expression pattern for the transcription factor *Zfp42* (*Rex1*). Only *Rex1*⁺ cells can efficiently colonize the blastocyst and contribute to embryonic development, a characteristic feature of ESCs in the naïve state of pluripotency (Toyooka et al., 2008). Therefore, a question arised as to whether *Rex1*⁺ and *Rex1*[–] ESCs differed in their cell cycle distributions. To address this issue, we took advantage of the *Rex1*^{GFP-*bla*}/*Oct4*^{CFP-*puro*} ESC line, which harbors a knock-in of the Green Fluorescent Protein (GFP) and *bla*^r selectable genes into *Rex1*, and a knock-in of the Cyan Fluorescent Protein (CFP) and *puro*^r selectable genes into *Oct4*. *Rex1*^{GFP-*bla*}/*Oct4*^{CFP-*puro*} ESCs were further engineered by lentiviral vector infection to express the orange-emitting Cdt1-mKO2 G1-specific fusion protein. The resulting cell line, *Rex1*^{GFP-*bla*}/*Oct4*^{CFP-*puro*}/*mKO2*, was continuously cultured in puromycin to eliminate *Oct4*[–] [CFP(–)] cells (Fig. 2A). Ninety percent of the CFP(+) cells exhibited a strong GFP fluorescence (GFP^{high}) and 10% exhibited a weak GFP fluorescence (GFP^{low}) (Figs. 2A,B). Real-time PCR analysis showed that the GFP^{low} and GFP^{high} phenotypes corresponded to *Rex1*[–] and *Rex1*⁺ cells, respectively. Both expressed *Oct4* at similar levels (Fig. 2C). We then determined the percentage of cells that expressed mKO2 in the two cell populations. mKO2(+) cells represented 15% of the GFP^{low} population and only 0.9% of the GFP^{high} population (Figs. 2B,D). Moreover, the mean intensity of mKO2-associated fluorescence was significantly higher in *Rex1*[–] cells than in *Rex1*⁺ cells (Fig. 2B). Together, these observations indicated that a larger fraction of the *Oct4*⁺/*Rex1*[–] ESC population was in the G1 phase compared to the *Oct4*⁺/*Rex1*⁺ population.

We next examined the changes in cell cycle distributions associated with the conversion of ESCs into EpiSCs as

paradigms of naïve and primed pluripotency. For this purpose, *Rex1*^{GFP-*bla*}/*Oct4*^{CFP-*puro*}/*mKO2* ESCs were treated with FGF2, activin, and fibronectin (Takehara et al., 2011). After 4 weeks (7 passages), the resulting cEpiSCs displayed the cardinal markers of primed pluripotency, including increased ectodermal (*Fgf5*), mesodermal (*Eomes*, *Brachyury*), and endodermal (*GATA6*, *FoxA2*) markers, and the downregulated naïve pluripotency markers (*Gbx2*, *Klf4*, *PECAM1*, and *Rex1*) (Supplementary data 2). An SSEA1⁺ sub-population was FACS-sorted from both ESCs and cEpiSCs, and the percentages of mKO2(+) cells in each were determined (Fig. 2E). SSEA1⁺ cEpiSCs had a dramatically higher percentage of mKO2(+) cells (25%) as compared to SSEA1⁺ ESCs (2.7%). A similar increase in the G1 fraction was observed in the *Oct4*⁺ sub-population (17.5% in cEpiSCs vs. 7.8% in ESCs). This indicated that the ESC-to-EpiSC transition was paralleled by an increase in the G1 phase duration. The percentage of mKO2(+) cells decreased in the SSEA1⁺ population to 0.7% when *Rex1*^{GFP-*bla*}/*Oct4*^{CFP-*puro*}/*mKO2* cells were cultured in N2B27 medium supplemented with MEK (PD0325901) and GSK3β (CHIR99021) inhibitors, which results in ESC self-renewal in the ground state of pluripotency (2i cells). Only rare mKO2(+) cells were observed in the GFP(–) population (Supplementary data 3).

Together, all of these results indicate that a short G1 phase is a characteristic of the ground and naïve pluripotent states, and that spontaneous or induced transition from the naïve state toward primed pluripotency is associated with a lengthening of the G1 phase.

mESCs in the G1 phase have an increased susceptibility to differentiation

We used the ESC–FUCCI line to determine whether the susceptibility of ESCs to retinoic acid (RA)-induced differentiation varied according to their position in the cell cycle. ESC–FUCCI were FACS-sorted into three distinct cell populations that corresponded to G1 [R1: mKO2(–) mAG(–) + mKO2(+) mAG(–)], S [R2: mKO2(–) mAG(+)], and G2 [R3: mKO2(–) mAG(++)] cell cycle phases (Fig. 3A). FACS-sorted cells were treated with 10^{–6} and 10^{–5} M RA (or with DMSO alone) for 1 h, cultured for 24 and 48 h in the absence of LIF, and subsequently restimulated with LIF for 5–7 days. This self-renewal assay determines the capacity of ESCs to withstand transient LIF starvation and subsequently, to form alkaline phosphatase-positive (AP⁺) undifferentiated colonies when plated at clonal density. The percentages of undifferentiated (AP⁺), mixed (AP⁺/AP[–]), and differentiated (AP[–]) colonies were determined (Figs. 3B,C, and Supplementary data 4). The percentages of mixed and differentiated colonies were significantly increased when cells were treated with 10^{–6} M RA during the G1 phase, as compared to cells that were treated during the other cell cycle phases. At 10^{–5} M, RA treatment during any cell cycle phase resulted in considerably increased percentages of these colonies. Moreover, the effect was more pronounced when cells were treated during the G1 phase. Of note, even without RA treatment, the G1 phase cells produced significantly more mixed and differentiated colonies as compared to the S and G2/M phase cells, which suggested that cells had an increased sensitivity to differentiation when they were transiting through the G1 phase prior to FACS sorting and replating at a low density.

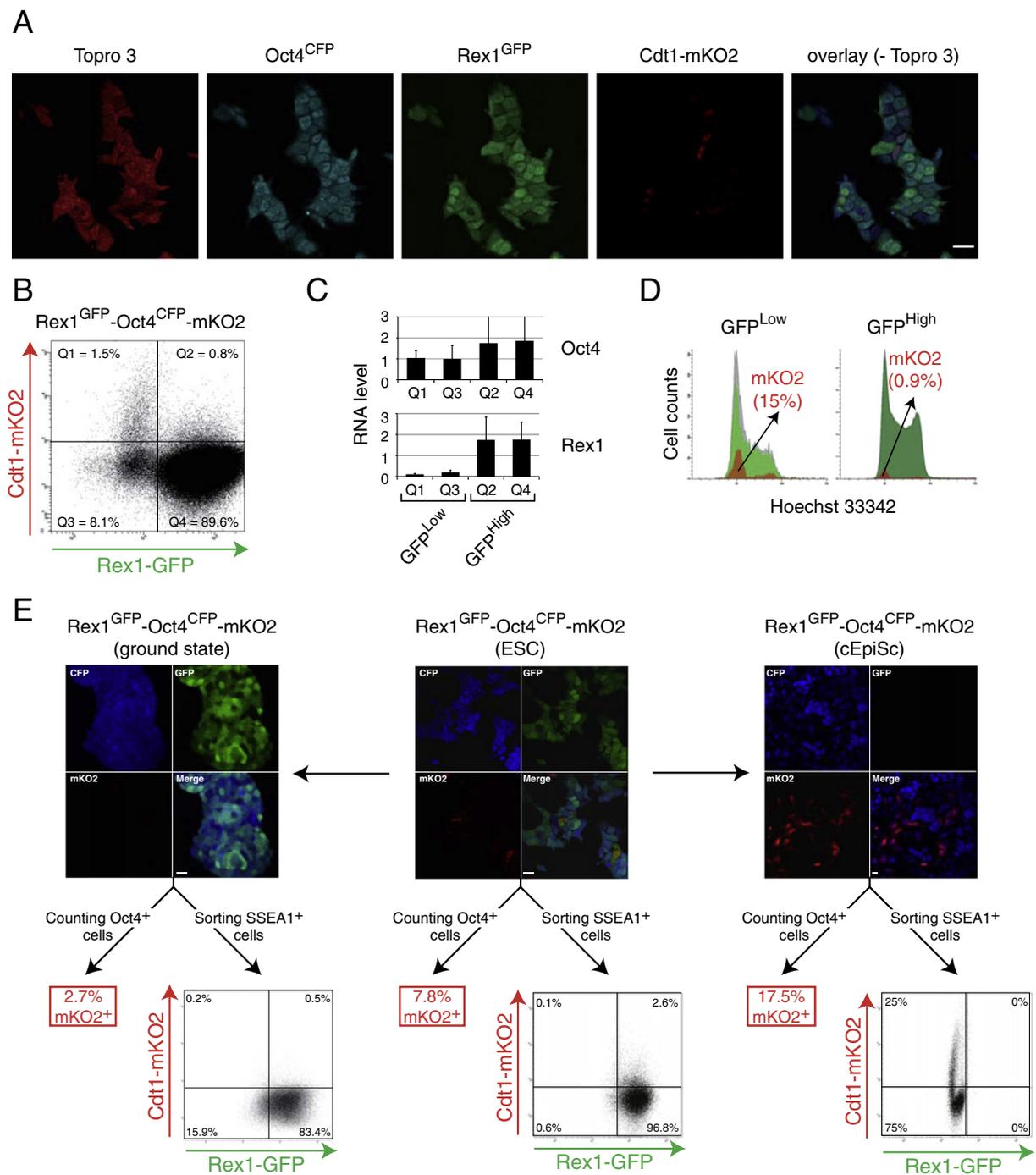


Figure 2 G1 phase duration varies according to the pluripotent state. (A) Representative fluorescence images of undifferentiated *Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2* ESCs that expressed CFP (blue), GFP (green), and mKO2 (red). (B) Dot plot of *Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2* ESCs showing GFP fluorescence associated with Rex1 expression and red fluorescence associated with mKO2 expression, analyzed by flow cytometry. (C) Histograms for the mRNA levels of *Oct4* and *Rex1* analyzed by real-time PCR in each fraction. (D) Histograms showing the cell cycle positions of the GFP^{low}/mKO2(+), GFP^{high}/mKO2(+), GFP^{low}/mKO2(-), and GFP^{high}/mKO2(-) fractions after staining with Hoechst 33342. (E) Upper panels: representative fluorescence images of *Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2* ESCs cultured in conditions with 2i/LIF (left panel), conditions with LIF + FCS (central panel), and after conversion into cEpiSCs (right panel). Bottom panels: dot plot of SSEA1⁺ *Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2* cells showing GFP fluorescence associated with Rex1 expression and red fluorescence associated with mKO2 expression, analyzed by flow cytometry. Scale bar represents 10 μ m.

Because $Rex1^-$ cells appeared to have a significantly longer G1 phase as compared to $Rex1^+$ cells, the increased sensitivity of the G1 phase cells to RA might have resulted

from the accumulation of $Rex1^-$ cells. To eliminate this confounding effect of cell population heterogeneity, we sought to eliminate $Rex1^-$ cells from the population prior to

the RA assay. For this purpose, *Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2* were sorted into four distinct cell populations that corresponded to S/G2/M *Rex1⁻* cells [P1: mKO2(-)GFP^{low}], G1 *Rex1⁻* cells [P2: mKO2(+)GFP^{low}], S/G2/M *Rex1⁺* cells [P3: mKO2(-)GFP^{high}], and G1 *Rex1⁺* cells [P4: mKO2(+)GFP^{high}] (Fig. 3D). The sorted cells were treated with 10⁻⁶ M RA for 1 h and subsequently analyzed with the LIF rescue assay. The percentages of mixed and differentiated colonies were significantly increased when GFP^{high} cells were treated with 10⁻⁶ M RA during the G1 phase as compared to similar cells treated during the other cell cycle phases (Figs. 3E,F). Moreover, the clonogenicity of the G1 phase GFP^{high} cells was significantly reduced ($p < 0.001$) as compared with the GFP^{high} cells in the other cell cycle phases (Fig. 3G). The same held true for GFP^{low} cells (*Rex1⁻*), which exhibited both an increased sensitivity to 10⁻⁶ M RA and a reduced clonogenicity while transiting through the G1 phase as compared with the other cell cycle phases. Collectively, these results indicated that *Rex1⁺* naïve pluripotent stem cells became highly susceptible to commitment to differentiation when transiting through the G1 phase.

Cyclin E-mediated alteration of the G1/S transition alters the balance between self-renewal and differentiation

Overexpression of cyclin E, encoded by the *CycE* gene, activity causes cells to progress through the G1 phase more rapidly and prematurely enter S-phase (Ohtsubo and Roberts, 1993; Resnitzky et al., 1994). Previous results have clearly shown that cyclin E-Cdk2 inhibition delays S phase entry (Ohtsubo et al., 1995; Tsai et al., 1993; van den Heuvel and Harlow, 1993). Based on our observations that the G1 phase provided a window of increased sensitivity to differentiation inducers, we assumed that overexpression of cyclin E would result in reinforced self-renewal, whereas knockdown of cyclin E would facilitate differentiation. To address this issue, ESCs were co-electroporated with the following plasmids: *CAG-irtetR-VP16-GBD*-IRES-puro*, a plasmid expressing a dexamethasone (Dex)-dependent rtTA (Anastassiadis et al., 2011); *pSport1-tetCMVratCycE^{WT}pA-INS-Hygro* [expressing a rat wild-type (WT) cyclin E cDNA]; *pSport1-tetCMVratCycE^{S180D}pA-INS-Hygro* (expressing a mutant form of rat cyclin E

with a S180D mutation that abolishes binding of cyclin E to Cdk2 (Matsumoto and Maller, 2004); and *pSport1-tetCMVhumCycE^{WT}pA-INS-Hygro* (expressing a human WT cyclin E cDNA). Each of these was under the control of the doxycyclin (Dox) inducible promoter *tetCMV*. Stably transfected cells were selected with appropriate antibiotics, and dually resistant clones were analyzed by real-time PCR to determine Dox/Dex-induced expression of WT or mutant cyclin E. Two clones, ESC-IND_ratCycE^{WT} and ESC-IND_ratCycE^{S180D}, exhibited a 60- and 20-fold increase in *CycE* RNA levels, respectively, as determined by qPCR using rat *CycE*-specific primers (Figs. 4A,B). They exhibited a 2.5- and 2-fold increase in *CycE* RNA levels, respectively, using primers that matched both mouse and rat *CycE*. The third clone, ESC-IND_humCycE^{WT}, exhibited an 18-fold increase in human *Cyclin E* RNA levels, as determined by qPCR using human *CycE*-specific primers. Robust expression of human cyclin E after treatment with Dox/Dex was confirmed by immunostaining (Fig. 4C). All three clones were analyzed with a LIF rescue assay. Dox/Dex-treated ESC-IND_ratCycE^{WT} produced a significantly higher percentage of undifferentiated colonies as compared to untreated cells, both in the presence of LIF and after LIF deprivation for 12, 24, 36, and 48 h (Fig. 4D). Dox/Dex-treated ESC-IND_humCycE^{WT} produced similar results. Of note, the overexpression of human cyclin E reduced the percentage of differentiated colonies by 2.5-fold upon LIF starvation for 48 h. In contrast, Dox/Dex-treated and -untreated ESC-IND_ratCycE^{S180D} exhibited no significant differences in their yields of undifferentiated colonies at all time points analyzed. Together, these results indicated that overexpression of cyclin E resulted in harnessing self-renewal and delaying the commitment to differentiation.

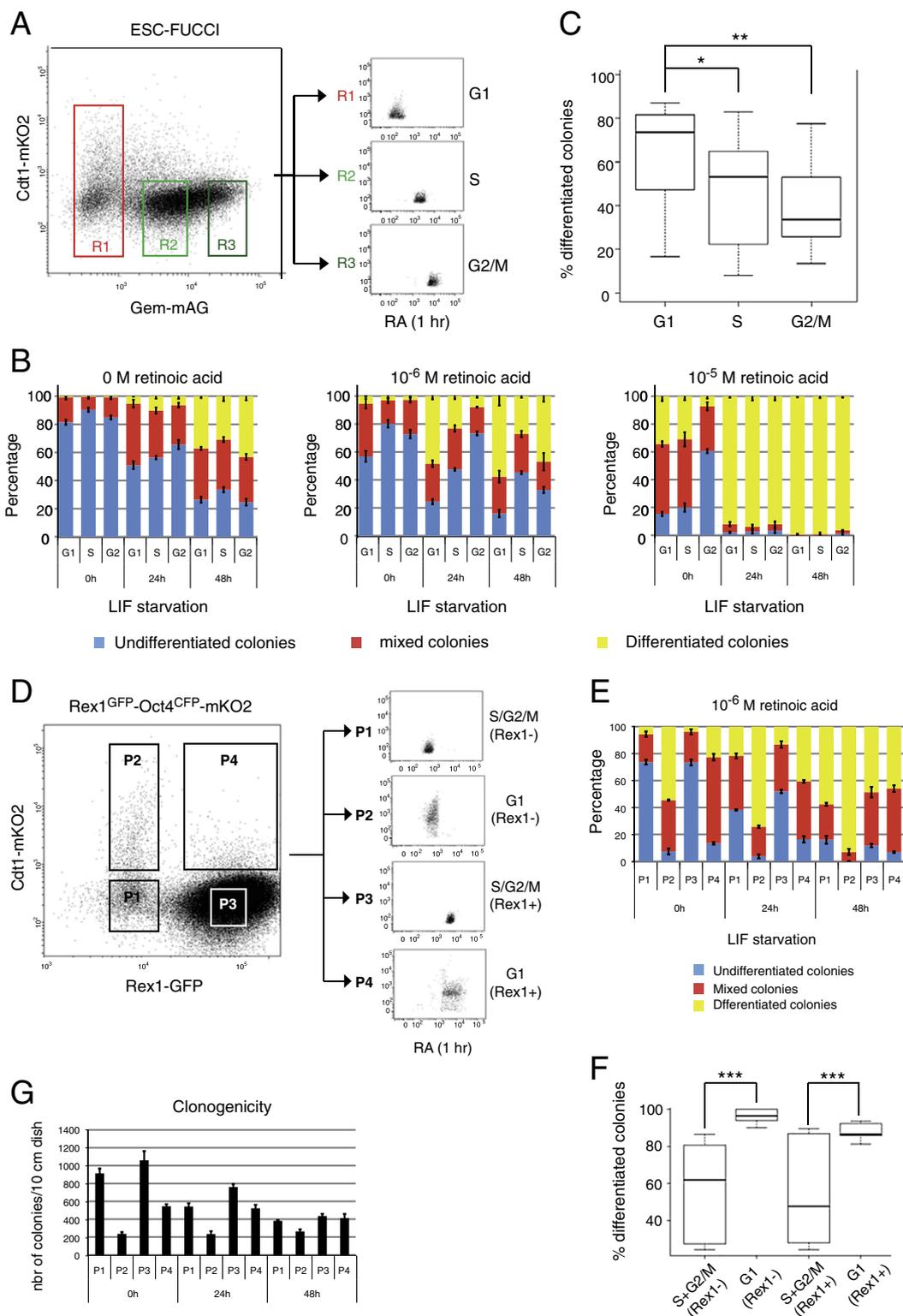
To further evaluate the role of cyclin E in the stabilization of the undifferentiated state, we knocked down cyclin E expression in ESC-FUCCI, by means of an interfering lentiviral vector expressing a *CycE* shRNA. A 5-fold reduction in *CycE* mRNA level, as compared to cells infected with the control vector (sh-scrambled), was observed after selection with G418 to kill the non-infected cells (Fig. 4E). ESC-FUCCI-shCycE cells formed smaller colonies, which could be explained by an increase in the G1 phase duration (26% of mKO2(+) cells in ESC-FUCCI-shCycE cells vs. 17% in control cells) (Fig. 4E).

When ESC-FUCCI-shCycE were induced to differentiate by EB formation, we observed both the reduced expression of

Figure 3 Cell-cycle dependency on RA-induced differentiation. (A) FACS sorting of ESC-Fucci in three distinct fractions, R1 [mKO2(-)mAG(-)+mKO2(+)mAG(-)], R2 [mKO2(-)mAG(+)], and R3 [mKO2(-)mAG(++)], corresponding to cells in the G1, S, and G2/M phases, respectively. (B) Histogram for the percentages of undifferentiated (blue), mixed (red), and differentiated (yellow) colonies observed in a LIF rescue assay after treatment with 0, 10⁻⁶, and 10⁻⁵ M retinoic acid for 1 h, followed by plating on gelatin-coated dishes at a density of 100 cells/cm². The duration of LIF starvation varied from 0 to 48 h, as indicated. (D) FACS sorting of *Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2* ESCs in four distinct fractions, P1 [GFP^{low}/mKO2(-)], P2 [GFP^{low}/mKO2(+)], P3 [GFP^{high}/mKO2(-)], and P4 [GFP^{high}/mKO2(+)], corresponding to *Rex1⁻/S-G2-M*, *Rex1⁻/G1*, *Rex1⁺/S-G2-M*, and *Rex1⁺/G1* cells, respectively. (E) Histogram for the percentages of undifferentiated (blue), mixed (red), and differentiated (yellow) colonies observed in a LIF rescue assay after treatment with 10⁻⁶ M retinoic acid for 1 h, followed by plating on gelatin-coated dishes at a density of 100 cells/cm². The duration of LIF starvation varied from 0 to 48 h, as indicated. (G) Histogram for the clonogenicity of each fraction after plating at a density of 100 cells/cm². (C,F) Box-and-whiskers plot showing the median (line inside the box) and the first and third quartiles (lower and upper edges of the box); the whiskers indicate the lowest and highest values within 1.5 times the interquartile range. Because the box plots limits overlapped and to confirm the tendency observed, we used a general linear model to quantify the effects of cell cycle phase on the percentages of differentiated colonies. *, $p < 0.05$; **, $p < 0.01$; ***, $p < 0.001$. (B,E,G) Means and standard errors were determined from three replicates.

pluripotency markers *Oct4* and *Nanog* and the advanced up-regulation of the endodermal markers *GATA4*, *GATA6* and *Sox17*, as compared to control cells (Fig. 4F). Notably, the ectodermal markers *Fgf5* and *Nestin* were expressed at lower levels in EBs made from ESC-FUCCI-shCycE, suggesting that

knockdown of cyclin E expression induced differentiation toward extra-embryonic endoderm at the expense of ectodermal derivatives. Together, these results indicate that the cyclin E level is rate-limiting during the G1-to-S phase transition and opposes the differentiation of naïve ESCs.



Lengthening of the G1 phase is an early, reversible response to LIF withdrawal

Withdrawing LIF from the ESC culture medium results in the lengthening of the G1 phase. Whether this process takes place before or after irreversible commitment to differentiation is an important issue for understanding the functional relationship between cell cycle regulation and self-renewal in ESCs. To address this issue, ESC–FUCCI were LIF-deprived for 24, 36, and 48 h, and then restimulated with LIF for 12, 24, 36, and 48 h. The percentages of G1 phase ESCs [mKO2(+)mAG(-) phenotype] increased by 1.23-, 1.74-, and 2.27-fold upon LIF deprivation for 24, 36, and 48 h, respectively (Figs. 5A,B). After stimulation with LIF for 12 h, these percentages further increased by 2.2-, 1.56-, and 1.23-fold (after LIF starvation for 24, 36, and 48 h, respectively), and then progressively returned to their original levels after 48 h of stimulation. The observed 12-h delay to achieve a maximum lengthening of the G1 phase might have reflected the duration of the entire cell cycle, as each cell had to pass through the G1 phase before undergoing cell cycle acceleration. Of note, after LIF starvation for 36 and 48 h, followed by restimulation for 48 h, many cells continued to accumulate mKO2 protein and went off-scale on the contour plots. These were most likely cells that were irreversibly committed to differentiation, and therefore failed to re-accelerate G1 phase transit upon LIF restimulation. This interpretation was supported by the observation that following LIF starvation for 36 and 48 h and despite restimulation with LIF, an increased number of cells exhibited a [mKO2(+)mAG(+)] phenotype that characterized differentiating cells (see Fig. 1D). The transient lengthening of the G1 phase was paralleled by transient alterations in gene expression patterns; specifically, there were decreased expressions of pluripotency markers (*Oct4*, *Nanog*, and *Rex1*) and increased expressions of lineage-specific markers (*Brachyury*, *Fgf5*, and *sox17*) (Fig. 5C). Overall, these markers returned to their original levels after LIF stimulation. However, after 36 and 48 h of LIF starvation followed by restimulation for 48 h, the expressions of *Brachyury*, *Fgf5*, and *sox17* did not return to their original levels, which indicated that many cells had been irreversibly committed to differentiation. Based on these results, we concluded that lengthening of the G1 phase is an early, reversible response to LIF starvation that takes place prior to commitment to differentiation.

LIF regulates G1 phase progression in the Rex1⁺ naïve cell population

The observed changes in G1 phase duration following LIF starvation might have resulted from a transient increase in the percentage of Rex1⁻ cells in the population. Therefore, to eliminate the confounding effect of varying Rex1⁺/Rex1⁻ ratios, we used the *Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2* ESC line to study the cell cycle response of Rex1⁺ ESCs to LIF stimulation. *Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2* cells were LIF-deprived for 24 h, and then restimulated with LIF for 12, 24, 36, and 48 h (Fig. 6A). LIF starvation resulted in a 4.4-fold increase in the percentage of Rex1⁻ cells, which returned to normal after LIF stimulation for 48 h (Fig. 6B). Consistent with the data obtained with ESC–FUCCI, the percentage of Rex1⁺ ESCs with a [mKO2(+)] phenotype increased by 2.8-fold upon LIF

deprivation for 24 h, and then returned to its original level after 24 h of LIF restimulation (Fig. 6C). In contrast, the percentage of Rex1⁻ ESCs that expressed a [mKO2(+)] phenotype only increased by 1.3-fold after LIF deprivation. These results indicated that stimulation with LIF signaling resulted in a considerable decrease in the G1 phase duration in the Rex1⁺ ESC sub-population, and to a much lesser extent in the Rex1⁻ ESC sub-population.

In order to match the observed changes in G1 phase duration with alterations in gene expression patterns in the Rex1⁺ cell population, the GFP^{high} population (comprising mKO2-positive and -negative cells) was sorted by FACS, and the expressions of pluripotency and lineage-specific markers were analyzed by quantitative RT-PCR (Figs. 6D,E). Only *Nanog* and *Fgf5* exhibited significant changes in their expression levels among the six genes that were tested. In contrast, the levels of expressions of *Oct4*, *Brachyury*, and *Sox17* remained constant. Thus, our results indicate that lengthening of the G1 phase is an early response to LIF withdrawal that takes place either shortly before or concomitantly with the transient activation of early lineage-specific markers in the Oct4⁺/Rex1⁺ naïve cell population.

Discussion

Our results show that a short G1 phase duration results from the activity of LIF signaling and that this short duration contributes to the maintenance of the naïve pluripotent state. We have presented the following five independent lines of evidence to support these conclusions: (i) the G1 phase duration increases as ESCs transit from the naïve pluripotent state to the primed state; (ii) the G1 phase of naïve ESCs is a phase of increased susceptibility to differentiation; (iii) overexpression of the G1-to-S phase transition-promoting cyclin E1 enhances self-renewal, whereas knocking down endogenous cyclin E1 expression results in lengthening of the G1 phase and spontaneous differentiation; and (iv) LIF signaling regulates the G1 phase duration in naïve ESCs.

FUCCI expression reveals somatic-like activities of APC^{cdh1} and SCF^{skp2} ubiquitination complexes in ESCs

It has been reported that the levels of major APC/C substrates, including Geminin, remain nearly constant during the pluripotent cell cycle (Fujii-Yamamoto et al., 2005; Yang et al., 2011). Our findings that the Geminin-mAG fusion protein was only expressed during the S and G2 phases and disappeared during the M and G1 phases, thus following a somatic-like pattern of expression, indicate that these previous conclusions were incorrect. A recent study had already questioned these conclusions by presenting a finding of the loss of Geminin expression during G1 transit in synchronized ESCs (Ballabeni et al., 2011). Our data corroborates this finding, using an experimental paradigm that does not require cell synchronization. We also showed that mKO2 expression was observed only in ESCs with a 2n DNA content. This was rapidly degraded during the G1-to-S phase transition, which demonstrated that SCF^{skp2} ubiquitination complexes were active throughout the S and G2/M phases. The Cdt1-mKO2 fusion protein is expressed at low levels in naïve ESCs. These levels moderately increase

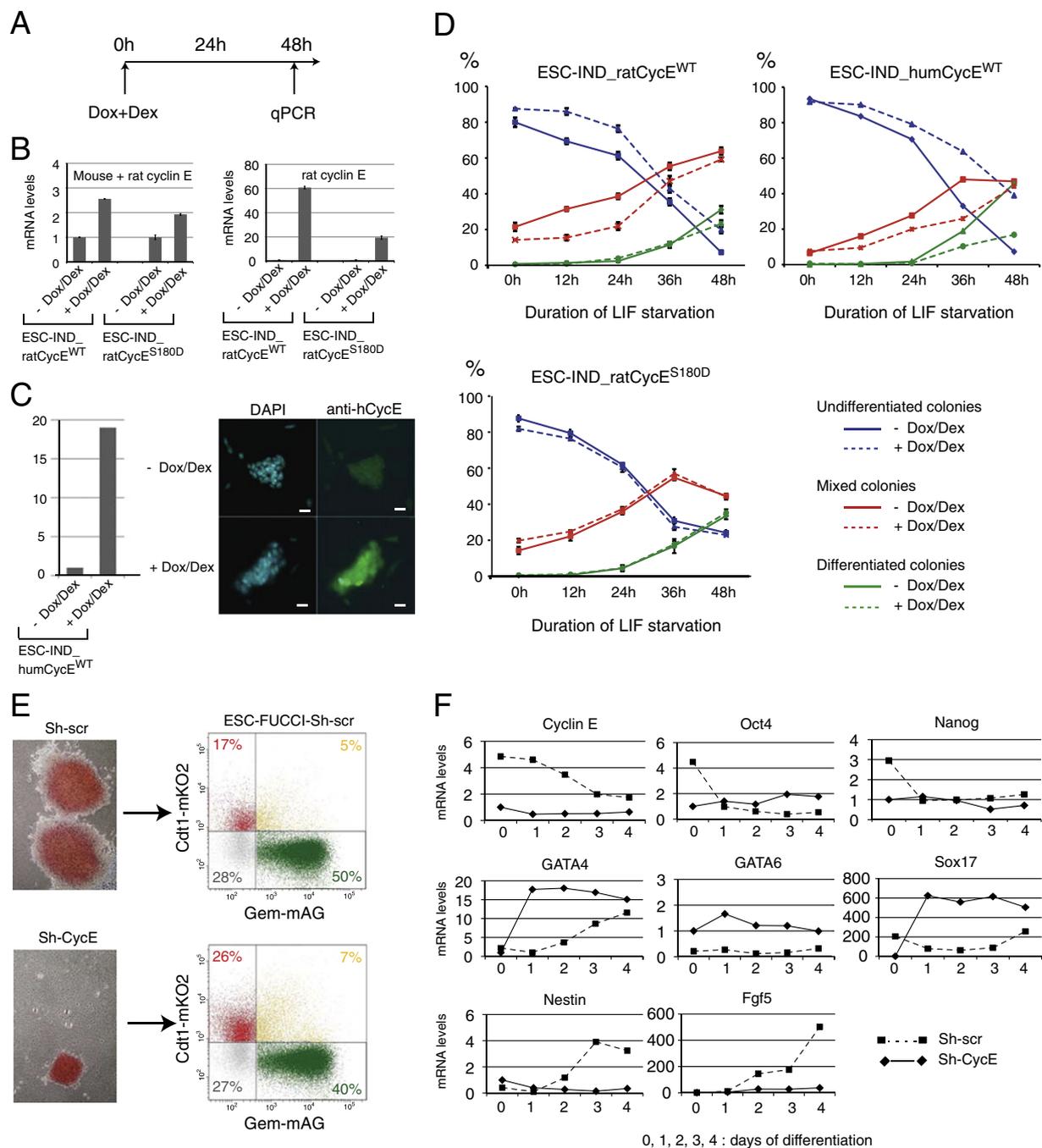


Figure 4 Cyclin E1 opposes differentiation. (A) Strategy of validation of the ESC-IND_Cyle1 cell lines. (B) qPCR analysis of *CycE* mRNA levels in ESC-IND_ratCyle1^{WT} (mouse/rat *CycE*-specific and myc-tagged *CycE*-specific primers), and ESC-IND_ratCyle1^{S180D} (mouse/rat *CycE*-specific and myc-tagged *CycE*-specific primers), before and after treatment with Dox/Dex. (C) Left panel: qPCR analysis of *CycE* mRNA levels in ESC-IND_humCyle1^{WT} (human *CycE*-specific primers). Right panel: immunostaining of human Cyclin E in ESC-IND_humCyle1^{WT}, using an antibody specific of human cyclin E. Scale bar represents 10 μ M. (D) LIF rescue assay with ESC-IND_ratCyle1^{WT}, ESC-IND_ratCyle1^{S180D}, and ESC-IND_humCyle1^{WT}. Means and standard errors were determined from three replicates. (E) Left panels: Morphology of ESC-Fucci-shCycE and ESC-FUCCI-shScrambled after staining for alkaline phosphatase activity. Right panels: Dot plots showing red [mKO2(+)/mAG(-)], yellow [mKO2(+)/mAG(+)], and green [mKO2(-)/mAG(+)] fluorescence. Figures indicate the percentages of cells of each color phenotype. (F) qPCR analysis for pluripotency and lineage-specific markers in ESC-Fucci-shCycE and ESC-FUCCI-shScrambled during differentiation (day 0 to day 4) induced by the formation of embryoid bodies.

as naïve cells convert to the primed pluripotent state and dramatically increase when they differentiate upon LIF withdrawal. We can explain these increases in mKO2-

associated fluorescence by the accumulation of the red fluorescence probe during G1 phase progression. Thus, the longer the G1 phase, the stronger is the mKO2-associated

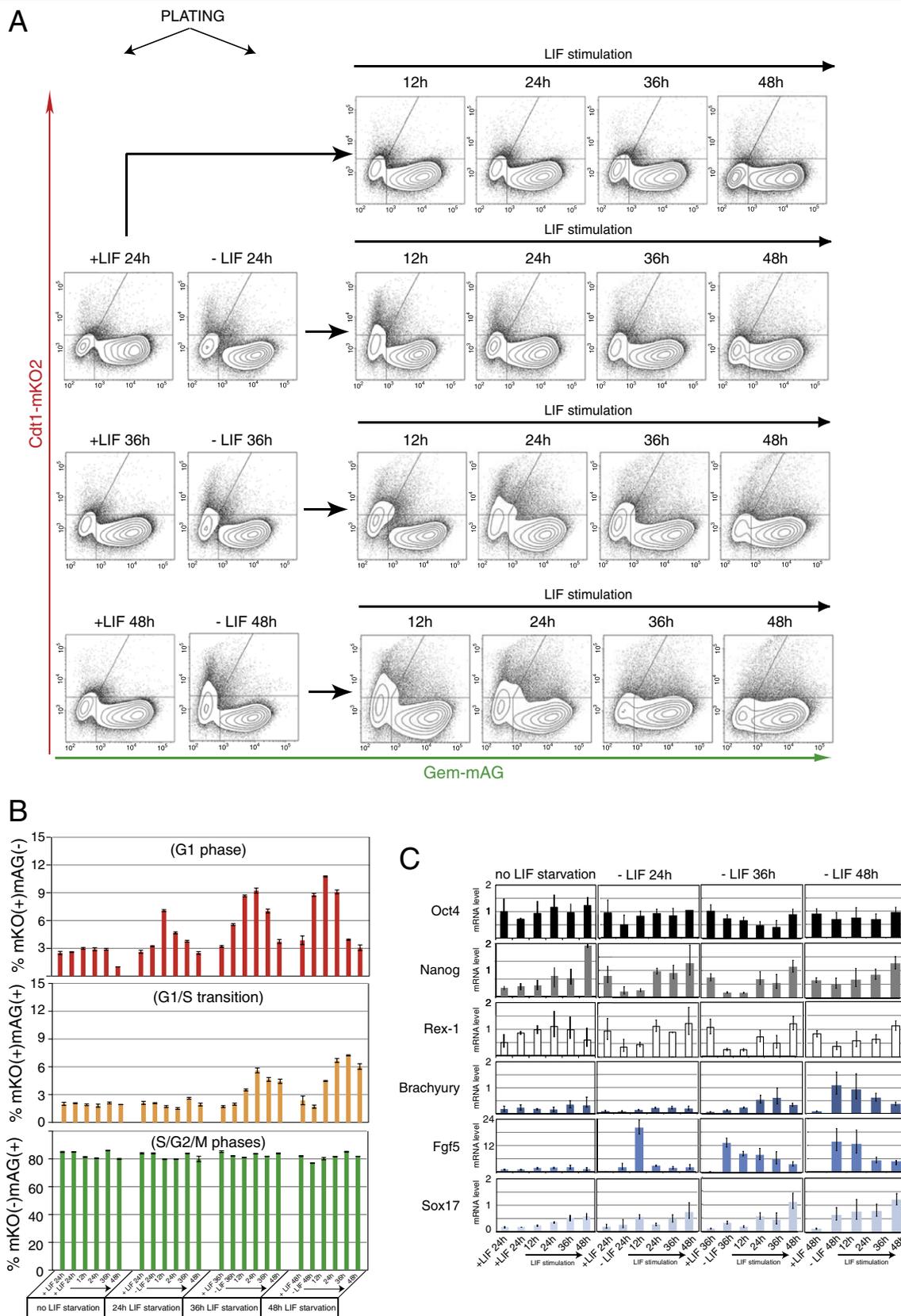


Figure 5 Lengthening of the G1 phase is an early, reversible response to LIF withdrawal. (A) Contour plots of ESC-FUCCI showing the fluorescence levels associated with mAG and mKO2 after withdrawing LIF for 24 h, followed by restimulation for 12, 24, 36, and 48 h. (B) Histograms for the percentages of mKO(+)/mAG(-) (G1), mKO(+)/mAG(+) (G1/S transition), and mKO(-)/mAG(+) (S-G2/M) cells in each of the experimental conditions defined in A. (C) qPCR analysis of pluripotency and lineage-specific markers after LIF starvation for 24 (-LIF 24 h), 36 (-LIF 36 h), and 48 (-LIF 48 h) h, followed by restimulation for 12, 24, 36, and 48 h. (B, C) “no LIF starvation” indicates control cells. Means and standard errors were determined from three replicates.

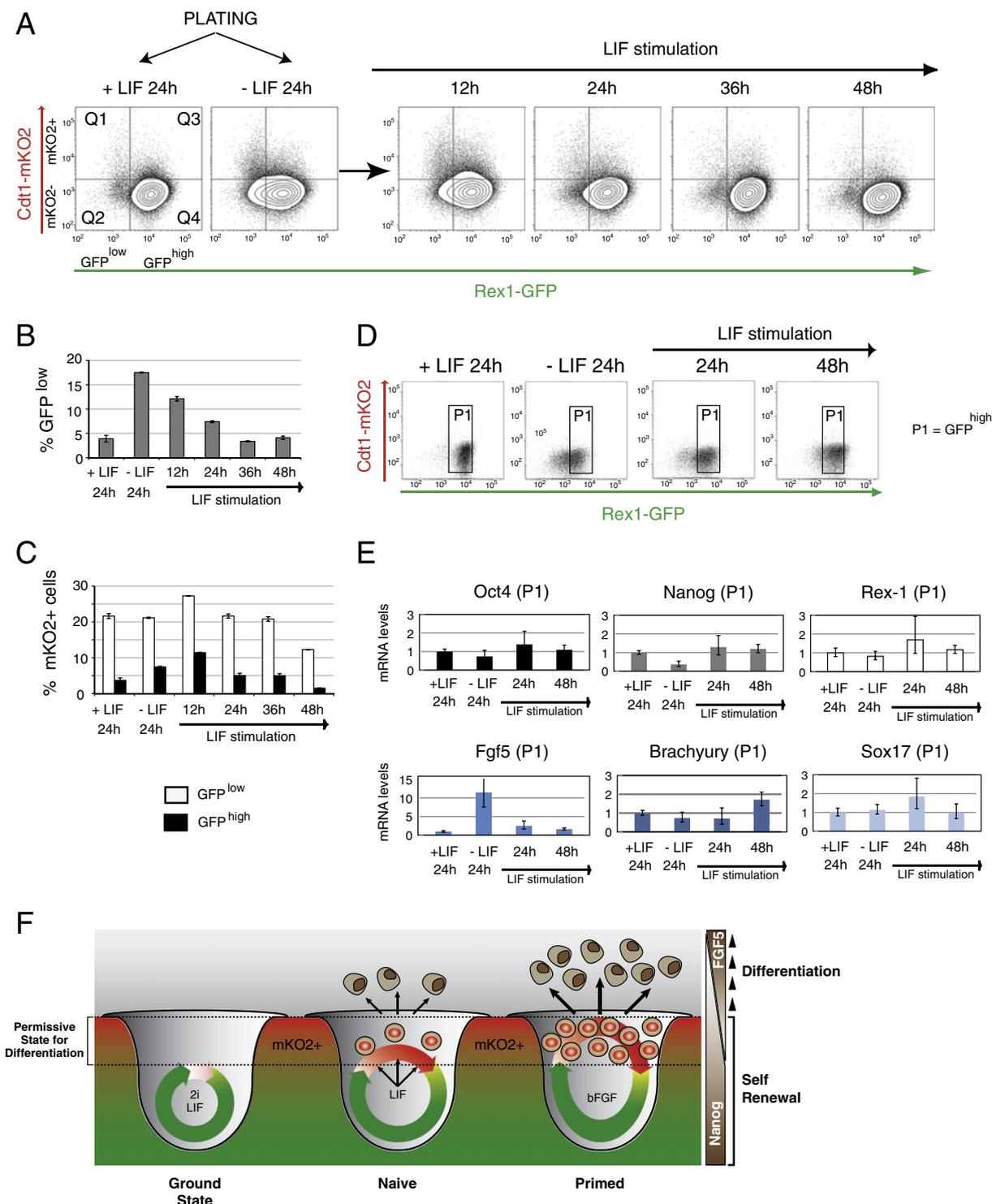


Figure 6 LIF regulates G1 phase progression in the Rex1⁺ cell population. (A) Contour plots of Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2 ESCs showing the fluorescence levels associated with GFP (Rex1 expression) and mKO2 after LIF withdrawal for 24 h, and followed by re-stimulation for 12, 24, 36, and 48 h. (B) Histogram for the percentages of GFP^{low} (Rex1⁻) cells [GFP^{low}/mKO2(-)] + [GFP^{low}/mKO2(+)] → [(Q1 + Q2)/(Q1 + Q2 + Q3 + Q4)]. (C) Histograms for the percentages of [GFP^{low}/mKO2(+)] [Rex1⁻ in G1 phase, Q1/(Q1 + Q2)] and [GFP^{high}/mKO2(+)] [Rex1⁺ in G1 phase, Q3/(Q3 + Q4)]. (D) FACS sorting of GFP^{high} cells from Rex1^{GFP-bla}/Oct4^{CFP-puro}/mKO2 ESCs treated as indicated. (E) Histograms for the mRNA levels of Oct4, Nanog, Rex1, Brachyury, Fgf5, and Sox17 in the P1 (GFP^{high}) fractions. (B, C, E) Means and standard errors were determined from three replicates. (F) Model for ESC self-renewal that incorporates the G1 phase for differentiation commitment and the role of LIF in regulating the G1 phase duration.

fluorescence. These increases could also be explained by Cdt1 stabilization during differentiation, independently from the G1 phase duration. However, we think that this is highly unlikely because it was recently reported that Cdt1 was actually more stable in mouse ESCs than in their differentiated derivatives (Ballabeni et al., 2011). Furthermore, the increase in mKO2-associated fluorescence was paralleled by an increase in the fraction of cells with a 2n DNA content, which further demonstrated the accumulation of cells in the G1 phase with respect to cells in the other cell cycle phases. Therefore, our data indicate that the FUCCI reporter system can be successfully used to study the cell cycle regulation of pluripotent stem cells in a wide variety of experimental paradigms.

ESCs exhibit an increased propensity to differentiate while they are transiting through the G1 phase

A link between the G1 phase and commitment of pluripotent stem cells into differentiation was first described in P19 embryonal carcinoma (EC) cells (Mummery et al., 1987). In ES cells, the same conclusion was proposed after observing that roscovitin, a pharmacological inhibitor of Cdk2, slowed down the mitotic cycle, delayed the G1-to-S phase transition, and induced spontaneous differentiation of mouse and human ESCs (Filipczyk et al., 2007; Koledova et al., 2009; Neganova et al., 2009). This differentiation, however, does not necessarily reflect an increased propensity to differentiate in the G1 phase. It may be caused by growth retardation irrespective of the cell cycle phase, or even by the direct effect of decreased Cdk2 activity on pluripotency regulators (Filipczyk et al., 2007). In a recent study, this issue was addressed more directly by separating live human ESCs at different cell cycle phases by elutriation, followed by analyzing their propensity to differentiate. It was shown that G1 cells were more prone to differentiate compared with S/G2 cells (Sela et al., 2012). However, this conclusion is flawed because spontaneously differentiating cells incorporate a longer G1 phase, and therefore would preferentially elutriate within the G1 fraction. Using the FUCCI reporter system, we could eliminate the spontaneously occurring Rex1⁻ cells and FACS-sort the G1 Oct4⁺/Rex1⁺ ESCs at nearly 100% purity. Hence, we could conclusively demonstrate that Oct4⁺ Rex1⁺ ESCs exhibited an increased propensity to differentiate during the G1 phase than the S and G2 phases. This conclusion was supported by our observation that overexpression of cyclin E increased the resistance of ESCs to transient LIF deprivation, whereas knockdown of cyclin E increased the G1 phase duration and facilitated differentiation. Thus, G1 provides a window of opportunity for differentiation signals, the aperture of which can be reduced or increased according to the level of kinase-active cyclin E. We also showed that the G1 phase duration varied according to the pluripotent state. G1 was shorter in 2i cells (ground state), and then gradually increased in duration as ESCs first progressed from the ground to the naïve state, and then from the naïve state to the primed state of pluripotency. It is tempting to speculate that this increase in G1 phase duration brings primed pluripotent stem cells closer to their commitment to differentiation. An important question is whether the increase in G1 phase duration that parallels the naïve-to-primed transition is associated with

changes in the mode of regulation of the G1-to-S phase transition.

LIF signaling contributes to the regulation of the G1 phase duration in naïve ESCs

Highlighting the role of LIF signaling in regulating the ESC cell cycle is difficult because of the confounding effect of differentiation, which can induce an increased G1 phase duration. To try to overcome this difficulty, we designed an experimental paradigm that distinguished the early effects of LIF starvation on the kinetics of G1 phase transit from its later effects on differentiation commitment. Using this paradigm, we observed that LIF withdrawal for 24 h reduced the kinetics of G1 phase transit of Oct4⁺/Rex1⁺ cells, and that LIF restimulation accelerated this transit so as to restore the original kinetics. This effect was observed prior to the irreversible changes in the expression levels of pluripotency and lineage-specific genes. We conclude that the altered kinetics of G1 transit induced by LIF starvation for 24 h is not an indirect consequence of differentiation commitment, but rather is the direct consequence of LIF-dependent regulation of the kinetics of G1 transit in Oct4⁺/Rex1⁺ ESCs. Thus, we propose that one function of LIF is to accelerate transit through the G1 phase and thereby protect ES cells from undesired differentiation signals. LIF withdrawal results in increasing this window of opportunity for differentiation cues (Fig. 6F). The molecular mechanism of this novel function of LIF remains to be explored.

Acknowledgments

This work was supported by the LabEX DEVweCAN (ANR-10-LABX-61). DC is a recipient of a fellowship from the Association pour la Recherche contre la Cancer (ARC). We are grateful to Peter Sicinski for the gift of the pCS2cycE^{WT} plasmid, and Jim Maller for the gift of pINDCycE^{WT} and pINDCycE^{S180D} plasmids. We are grateful to Sébastien Dussurget and Thibault Andrieu (IFR Bioscience-Gerland) for their assistance with flow cytometry.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <http://dx.doi.org/10.1016/j.scr.2012.10.004>.

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